In the Specification:

Please replace the paragraph at page 22, from line 23 through page 23 line 7, with the following paragraph:

-- The MPR1128 promoter (SEQ ID NO. 4) is derived from PrHMWG-Dx5 (SEQ ID NO. 1) by deleting the sequence located upstream of nucleotide –238, this sequence comprising the two prolamine-"like" boxes and the two GATA boxes. The promoter fragment was amplified by PCR from 5 ng of pMRT1125 matrix DNA (described in section 2.1 of Example 2) with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCCAGAACTAGGATTACGCCG 3' (SEQ ID NO. 35), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of the Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 30 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 50°C for 1 min. and elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min.--

Please replace the paragraph at page 23 line 25 through page 24, line 4, with the following paragraph:

-- The MPr1127 promoter (SEQ ID NO. 3) is derived from the HMWG-Dx5 promoter (SEQ ID NO. 1) by deleting the sequence located upstream of nucleotide –205, this sequence comprising the two prolamine-"like" boxes, the two GATA boxes and the "G" box. The promoter fragment was amplified by PCR and treated in the same way as the MPR1128 promoter (SEQ ID NO. 4)(described in section 2.2 of Example 2), except that the 2 oligodeoxynucleotides used are 5' ATCGGGAATTCGCAGACTGTCCAAAAATC 3' (SEQ ID NO. 15), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site.--

Please replace the paragraph at page 24, from line 8 through line 15, with the following paragraph:

--The MPr1126 promoter (SEQ ID NO. 2) is derived from the HMWG-Dx5 promoter (SEQ ID NO. 1) by deleting the sequence located upstream of nucleotide –142, this sequence comprising the two prolamine-"like" boxes, the two GATA boxes, the "G" box and the activating element. The promoter fragment was amplified by PCR and treated in the same way as the MPR1128 promoter (SEQ ID NO. 4)(described in section 2.2 of Example 2), except that the 2 oligodeoxynucleotides used are 5' ATCGGAATTCGTGTTGGCAAACTGC 3' (SEQ ID NO. 14), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3 (SEQ ID NO. 29), possessing the BamHI restriction site.--

Please replace the paragraph at page 24 from line 19 through line 28, with the following paragraph:

-- The MPr1183 promoter results from the insertion of an XbaI restriction site upstream of the MPR1128 promoter (SEQ ID NO. 4) (described in section 2.2 of Example 2). The promoter fragment was amplified by PCR from 5 ng of pMRT1128 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' TCggAATTCTAgACgCCgATTACgTggCTTTAgC 3 (SEQ ID NO. 37), containing the EcoRI and XbaI restriction sites, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 30 cycles each consisting of the steps of denaturation at 94°C for--

Please replace the paragraph at page 25, from line 9 through line 23, with the following paragraph:

-- The ligation was carried out with 100 ng of the MPR1128 promoter fragment thus treated and 50 ng of plasmid pGem3Z-1 (described in section 2.1 of Example 2) overnight at

16°C in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs). *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCgCAgCCATggTCCTgAACC 3' (SEQ ID NO. 25) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), in the presence of 15 nmol of each of the dNTPs, of the Taq DNA polymerase buffer (1X), of 75 nmol of MgCl2 and of 1.25 U of Taq DNA polymerase (Promega Corp.) in a 50 µl reaction volume. The amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 3 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 30 sec., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. During the final cycle, the elongation was continued at 72°C for 5 min.--

Please replace the paragraph at page 26, line 25 through page 27, line 11, with the following paragraph:

-- It was constructed by fusing, at the NcoI restriction site of pMRT1183 (described in section 2.5 of Example 2), a fragment amplified by PCR from 5 ng of pMRT1128 matrix DNA with the aid of 100 pmol each of the 2 oligodeoxynucleotides 5' ATCggAATTCTAgACgCCgATTACgTggCTTTAgC 3' (SEQ ID NO. 37), containing the EcoRI and XbaI restriction sites, and 5' CATgCCATggCCAACACAAAAgAAgCTgg 3' (SEQ ID NO. 32), possessing the NcoI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 10 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The DNA fragment derived from the amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit, and hydrolysed successively with NcoI and XbaI, for 1 h at 37°C.--

Please replace the paragraph at page 28, from line 7 through line 22, with the following paragraph:

-- The "MPr1216 (SEQ ID NO. 21) 5' fragment", synthesized by PCR, was amplified from 5 ng of pMRT1128 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCgCCgATTACgTggCTTTAgC 3' (SEQ ID NO. 24), containing the EcoRI restriction site, and 5' gCTCTAgACCAACACAAAAgAAgCTgg 3' (SEQ ID NO. 31) possessing the XbaI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 10 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The DNA fragment derived from the PCR amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit, and hydrolysed with EcoRI for 1 h at 37°C. The DNA fragment was then subjected to the action of 20 U of the Klenow fragment (New England Biolabs) for 30 min at 37°C in the presence of 60 nmol of each of the dNTPs, of 12 µl of 500 mM Tris-HCL, pH 7.5/500 mM MgCl2 buffer and 6 µl of 1 M DTT, and digested with XbaI for 1 h at 37°C.--

Please replace the paragraph at page 28. line 26 through page29, line 8, with the following paragraph:

-- The ligation reaction was carried out with 50 ng of the "MPr1216 (SEQ ID NO.SEQ.ID21) 5' fragment" and 50ng of the "MPr1216 (SEQ ID NO. SEQ.ID21) 3' fragment" thus treated, and 50 ng of plasmid pGem3Z-1, in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs), in the "GeneAmp PCR System 9700" thermocycler, as described above. *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 10 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCgCCgATTACgTggCTTTAgC 3' (SEQ ID NO.

24) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29) in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 29, from line 16 through line 29, with the following paragraph:

-- The MPr1217 promoter (SEQ ID NO. 22) was constructed by inserting, into the XbaI restriction site of pMRT1183 (described in section 2.5 of Example 2), two identical promoter fragments synthesized by PCR from 5 ng of matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCTAgACgCCgATTACgTggCTTTAgC 3' (SEQ ID NO. 37), containing the EcoRI and XbaI restriction sites, and 5' gCTCTAgACCAACACAAAAgAagCTgg 3' (SEQ ID NO. 31), possessing the XbaI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 10 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The DNA fragment derived from the PCR amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit, and hydrolysed with XbaI for 1 h at 37°C.--

Please replace the paragraph at page 30, from line 5 through line 14, with the following paragraph:

-- The ligation reaction was carried out with 50 ng of promoter fragment and 100 ng of vector fragment thus prepared, in a 10 µl reaction mixture, in the presence of the 1X T4 DNA ligase buffer (New England Biolabs) and of 400 units of T4 DNA ligase (New England Biolabs) in the "GeneAmp PCR System 9700" thermocycler, as described above. *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 10 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCgCCgATTACgTggCTTTAgC 3' (SEQ ID NO.

24) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29) in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 30, line 27 through page 31, line 15, with the following paragraph:

-- The "MPr1130 (SEQ ID NO. SEQ.ID05) 5' fragment" was amplified by PCR from 5 ng of pUC19-HMWG matrix DNA (described in section 2.1 of Example 2) with the aid of 20 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 21), containing the EcoRI restriction site, and 5' TgCgTCATCCCTTACgTCAgTggAgATATCACATCAATCTTgATATCACATCAATCACggT gAggTTTgTTTAgCCTAAg 3' (SEQ ID NO. 34), possessing the 55-bp sequence corresponding to a duplication of the as-2 motif (Lam and Chua, 1989, *supra*) and to the as-1 motif (Lam et al., 1989, *supra*) of CaMV 35S, in the presence of 10 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs), in a 50 µl reaction volume. The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 15 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. Forty µl of the PCR reaction medium were then subjected to the action of 12.5 U of the Klenow fragment (New England Biolabs) in the presence of 20 nmol of each of the dNTPs for 10 min. at 37°C. The PCR product thus treated was then isolated on a 1.5% agarose gel with the aid of the "QIAquick Gel Extraction" kit.--

Please replace the paragraph at page 31, from line 16 through line 22, with the following paragraph:

-- The "MPr1130 (SEQ ID NO. 5) 3' fragment" was synthesized and treated in the same way as the "MPr1130 (SEQ ID NO. 5) 5' fragment", except that the 2 oligodeoxynucleotides used are 5' ATTgATgTgATATCAAgATTgATgTgATATCTCCACTgACgTAAgggATgACgCACACgCAg CCATggTCCTgAACCTTC 3' (SEQ ID NO. 27), possessing the 55-bp sequence corresponding to a duplication of the as-2 motif (Lam and Chua, 1989, *supra*) and to the as-1 motif (Lam et al.,

1989, *supra*) of CaMV 35S, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), containing the BamHI restriction site.

Please replace the paragraph at page31, from line 23 through page 32, line 11, with the following paragraph:

-- The "MPr1130 (SEQ ID NO. 5) 5' fragment" and the "MPr1130 (SEQ ID NO. 5) 3' fragment" were then assembled by overlap extension so as to generate the "MPr1130 fragment (SEQ ID NO. 5)". In order to do this, a PCR amplification was carried out using 7.5 µl of each of the two PCR products thus treated, with the aid of 20 pmol of each of the oligodeoxynucleotides 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 36), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site, in the presence of 10 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs), in a 50 µl reaction volume. The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 15 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The "MPr1130 fragment (SEQ.ID0SEQ ID NO. 5)" thus synthesized was isolated on a 1.5% agarose gel with the aid of the "QIAquick Gel Extraction" kit. This fragment was then hydrolysed with EcoRI for 1 h at 37°C, and subjected to the action of 20 U of the Klenow fragment (New England Biolabs) for 30 min at 37°C in the presence of 60 nmol of each of the dNTPs, of 12 µl of 500 mM Tris-HCL, pH 7.5/500 mM MgCl2 buffer and 6 of µl of 1 M DTT. Finally, the MPr1130 fragment (SEQ ID NO. SEQ.ID05) was digested with BamHI for 1 h at 37°C.--

Please replace the paragraph at page 32, from line 12 through line 21, with the following paragraph:

-- The ligation was carried out with 100 ng of the "MPr1130 fragment (SEQ ID NO.5)" thus treated and 50 ng of plasmid pGem3Z-1 (described in section 2.1 of Example 2) overnight

at 16°C in a 10 µl reaction mixture, in the presence of 1 µl of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs). *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 36) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29) in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 33, from line 3 through line 9, with the following paragraph:

-- The "MPr1131 (SEQ ID NO. 6) 5' fragment" was synthesized and treated in the same way as the "MPr1130 (SEQ ID NO. 5) 5' fragment" (described in section 3.1 of Example 3), except that the 2 oligodeoxynucleotides used are 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 36), containing the EcoRI restriction site, and 5' TgCgTCATCCCTTACgTCAgTggAgATATCACATCAATCACggTgAggTTTgTTTAgCCTAA g 3' (SEQ ID NO. 33), possessing the 38-bp sequence corresponding to the as-2 motif (Lam and Chua, 1989, *supra*) and to the as-1 motif (Lam et al., 1989, *supra*) of CaMV 35S.--

Please replace the paragraph at page 33, from line 10 through line 17, with the following paragraph:

-- The "MPr1131 (SEQ ID NO. 6) 3' fragment" was synthesized and treated in the same way as the "MPr1130 (SEQ ID NO. 5) 5' fragment" (described in section 3.1 of Example 3), except that the 2 oligodeoxynucleotides used are 5' ATTgATgTgATATCTCCACTgACgTAAgggATgACgCACACgCAgCCATggTCCTgAACCTT C 3' (SEQ ID NO. 27) possessing the 38-bp sequence corresponding to the as-2 motif (Lam and Chua, 1989, *supra*) and to the as-1 motif (Lam et al., 1989, *supra*) of CaMV 35S, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29) containing the BamHI restriction site.--

Please replace the paragraph at page 33, from line 18 through line 29, with the following paragraph:

--The "MPr1131 (SEQ ID NO. 6) 5' fragment" and the "MPr1131 (SEQ ID NO. 6) 3' fragment" were then assembled by overlap extension so as to generate the "MPr1131 fragment (SEQ ID NO. 6)". In order to do this, a PCR amplification was carried out using 7.5 µl of each of the two PCR products thus treated, with the aid of 20 pmol of each of the oligodeoxynucleotides 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 36), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site, in the presence of 10 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs), in a 50 µl reaction volume. The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 15 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation--

Please replace the paragraph at page 34, from line 8 through line 17, with the following paragraph:

-- The ligation was carried out with 100 ng of the "MPr1130 fragment (SEQ ID NO. 5)" thus treated and 50 ng of plasmid pGem3Z-1 (described in section 2.1 of Example 2) overnight at 16°C in a 10 µl reaction mixture, in the presence of 1 µl of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs). *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCCAgCTTTgAgTggCCgTAg 3' (SEQ ID NO. 36) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29) in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 34, from line 25 through page 35, line 6, with the following paragraph:

-- The promoter fragment was amplified by PCR from 5 ng of pMRT1130 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCCAGAACTAGGATTACGCCG 3' (SEQ ID NO. 35), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3' (SEQ ID NO. 29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of the Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 1 min.--

Please replace the paragraph at page 35, from line 13 through line 22, with the following paragraph:

-- The ligation was carried out with 100 ng of the MPR1135 promoter fragment (SEQ ID NO. 9) thus treated and 50 ng of plasmid pGEM3Z-1 (described in section 2.1 of Example 2) overnight at 16°C in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs). *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCGTGTTGGCAAACTGC 3' (SEQ ID NO. 14) and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3'(SEQ ID NO. 29), in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 36, from line 13 through line 18, with the following paragraph:

--The promoter fragment was amplified by PCR from 5 ng of pMRT1131 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGGAATTC-GCAGACTGTCCAAAAATC 3', (SEQ ID NO:15), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), possessing the BamHI

restriction site, treated and obtained in the same way as the MPr1135 promoter (SEQ ID NO. 9) (described in section 3.3 of Example 3). --

Please replace the paragraph at page 37, from line 1 through line 6, with the following paragraph:

--The promoter fragment was amplified by PCR from 5 ng of pMRT1130 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGGAATTCGC-AGACTGTCCAAAAATC 3', (SEQ ID NO:15), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), possessing the BamHI restriction site, treated and obtained in the same way as the MPr1135 promoter (SEQ ID NO. 9) (described in section 3.3 of Example 3).--

Please replace the paragraph at page 37, from line 14 through line 19, with the following paragraph:

--The promoter fragment was amplified by PCR from 5 ng of pMRT1131 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTC-GTGTTGGCAAACTGC 3', (SEQ ID NO:14), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), possessing the BamHI restriction site, treated and obtained in the same way as the MPr1135 promoter (SEQ ID NO. 9) (described in section 3.3 of Example 3).--

Please replace the paragraph at page 38, from line 1 through line 6, with the following paragraph:

--The promoter fragment was amplified by PCR from 5 ng of pMRT1130 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCGT-GTTGGCAAACTGC 3', (SEQ ID NO:14), containing the EcoRI restriction site, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), possessing the BamHI restriction site, treated and obtained in the same way as the MPr1135 promoter (SEQ ID NO. 9) (described in section 3.3 of Example 3).--

Please replace the paragraph at page 38, from line 15 through line 26, with the following paragraph:

--The MPr1139 promoter (SEQ ID NO. 13) was amplified by PCR from 5 ng of pMRT1131 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCTCgACATggTTAgAAgTTTTgAgTgCCgCCACTACTCgACAT-ggTTAgAAgTTTTgAgTggCCgTAgATTTgC 3', (SEQ ID NO:23), containing the EcoRI restriction site and the two "cereal" boxes described above, and 5' TACggATCCCCgggg-ATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of the Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 1 min.--

Please replace the paragraph at page 39, from line 5 through line 14, with the following paragraph:

--The ligation was carried out with 100 ng of the MPr1139 promoter fragment (SEQ ID NO. 13) thus treated and 50 ng of plasmid pGem3Z-1 (described in section 2.1 of Example 2) overnight at 16°C, in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs). *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCCAGAACTAGGATTACGCCG 3', (SEQ ID NO:35), and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please replace the paragraph at page 39, from line 25 through page 40, line 12, with the following paragraph:

-- -The "MPr1200 (SEQ ID NO. 19) 5' fragment", synthesized by PCR, was amplified from 5 ng of pMRT1139 matrix DNA (described in section 3.9 of Example 3) with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCTCgACATgg 3', (SEQ ID NO:26), containing the EcoRI restriction site, and 5' gCTCTAgAgCAAATCTACggCCACTC 3', (SEQ ID NO:30), possessing the XbaI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 10 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The DNA fragment derived from the PCR amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit, and hydrolysed with EcoRI for 1 h at 37°C. The DNA fragment was then subjected to the action of 20 U of the Klenow fragment (New England Biolabs) for 30 min at 37°C in the presence of 60 nmol of each of the dNTPs, of 12 µl of 500 mM Tris-HCL, pH 7.5/500 mM MgCl2 buffer and 6 µl of 1 M DTT, and then digested with XbaI for 1 h at 37°C.--

Please replace the paragraph at page 40, from line 13 through line 22, with the following paragraph:

-- -The "MPr1200 (SEQ ID NO. 19) 3' fragment", synthesized by PCR, as amplified from 5 ng of pMRT1138 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCTAgACgCCgATTACgTggCTTTAgC 3', (SEQ ID NO:37), containing the EcoRI restriction site and XbaI restriction site, and 5' TACggATCCCCggggATCTCTAgTTT-gTggTgC 3', (SEQ ID NO:29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs) in the "GeneAmp PCR System 9700" thermocycler, under

the same conditions as the "MPr1200 (SEQ ID NO. 19) 5' fragment". The DNA fragment derived from the PCR amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit and hydrolysed successively with XbaI and BamHI for 1 h at 37°C.--

Please replace the paragraph at page 40, from line 23 through page 41, line 3, with the following paragraph:

--The ligation reaction was carried out with 50 ng of the "MPr1200 (SEQ ID NO. 19) 5' fragment", 50 ng of the "MPr1200 (SEQ ID NO. 19) 3' fragment" thus treated, and 50 ng of plasmid pGem3Z-1, in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs) in the "GeneAmp PCR System 9700" thermocycler, as described above. *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 10 pmol of each of the 2 oligodeoxynucleotides 5' ATCggAATTCgCAgCCATggTCCTgAACC 3', (SEQ ID NO:25), and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), in the "GeneAmp PCR System 9700" thermocycler, as described above. --

Please replace the paragraph at page 41, from line 14 through page 42, line 2, with the following paragraph:

-- - The "MPr1213 (SEQ ID NO. 20) 5' fragment", synthesized by PCR, was amplified from 5 ng of pMRT1139 matrix DNA (described in section 3.9 of Example 3) with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCTCgACATgg 3', (SEQ ID NO:26), containing the EcoRI restriction site, and 5' gCTCTAgAgCAAATCTACggCCACTC 3', (SEQ ID NO:30), possessing the XbaI restriction site, in the presence of 50 nmol of each of the dNTPs, of Vent DNA polymerase buffer (1X) and of 2 U Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for

10 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 5 min. The DNA fragment derived from the PCR amplification was isolated on a 1.5% agarose gel with the aid of the "Concert Rapid Gel Extraction System" kit, and hydrolysed with EcoRI for 1 h at 37°C. The DNA fragment was then subjected to the action of 20 U of the Klenow fragment (New England Biolabs) for 30 min at 37°C in the presence of 60 nmol of each of the dNTPs, of 12 µl of 500 mM Tris-HCL, pH 7.5/500 mM MgCl2 buffer and 6 µl of 1 M DTT, and digested with XbaI for 1 h at 37°C.--

Please replace the paragraph at page 42, from line 7 through line 16, with the following paragraph:

--The ligation reaction was carried out with 50 ng of the "MPr1213 (SEQ ID NO. 20) 5' fragment" and 50 ng of the "MPr1213 (SEQ ID NO. 20) 3' fragment" thus treated, and 50 ng of plasmid pGem3Z-1, in a 10 µl reaction mixture, in the presence of the T4 DNA ligase buffer (1X) and of 400 units of T4 DNA ligase (New England Biolabs) in the "GeneAmp PCR System 9700" thermocycler, as described above. *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 10 pmol of each of the 2 oligodeoxynucleotides 5' TACgAATTCCTCgACATgg 3', (SEQ ID NO:26), and 5' gCTCTAgAgCAAATCTACggCCACTC 3', (SEQ ID NO:30),in the "GeneAmp PCR System 9700" thermocycler, as described above. --

Please replace the paragraph at page 42, from line 24 through page 43, line 6, with the following paragraph:

--The MPr1199 promoter (SEQ ID NO. 18) was amplified by PCR from 5 ng of pMRT1128 matrix DNA with the aid of 100 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCAAATGGGCCGGACCGGGCCGGCCCAGCGCCGATTACGTGGCT-TTAGC 3', (SEQ ID NO:38), containing the "GC-rich" element described above and the EcoRI and XbaI restriction sites, and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID

NO:29), possessing the BamHI restriction site, in the presence of 50 nmol of each of the dNTPs, of the Vent DNA polymerase buffer (1X) and of 2 U of Vent DNA polymerase (New England Biolabs). The PCR amplification reaction was carried out in the "GeneAmp PCR System 9700" thermocycler. After a denaturation at 94°C for 5 min., the DNA was subjected to 25 cycles each consisting of the steps of denaturation at 94°C for 1 min., of hybridization at 55°C for 1 min. and of elongation at 72°C for 1 min. 30 sec. During the final cycle, the elongation was continued at 72°C for 1 min.--

Please replace the paragraph at page 43, from line 13 through line 22, with the following paragraph:

--The ligation was carried out with 100 ng of the MPr1199 promoter fragment (SEQ ID NO. 18) thus treated and 50 ng of plasmid pGem3Z-1 (described in section 2.1 of Example 2), with PCR cycles in the "GeneAmp PCR System 9700" thermocycler under the conditions described above. *Escherichia coli* DH5a bacteria, which had been made competent beforehand, were transformed with all of the ligation reaction mixture. The plasmid DNA of the clones obtained, which were selected on LB medium supplemented with ampicillin (50 mg/l), was analysed by PCR with the aid of 25 pmol of each of the 2 oligodeoxynucleotides 5' ATCGGAATTCCAGAACTAGGATTACGCCG 3', (SEQ ID NO:35), and 5' TACggATCCCCggggATCTCTAgTTTgTggTgC 3', (SEQ ID NO:29), in the "GeneAmp PCR System 9700" thermocycler, as described above.--

Please delete the phrase, "What is claimed is:" from page 70.

Please insert the phrase, "What is claimed is" at page 71 after "**CLAIMS**" and before "1".